

KB03037
Phenolic Compounds
Assay Kit

96 well plate 100/200/400 tests





Table of contents

| ١. | General information | l |
|-----|--|----|
| 2. | Technical specifications | 2 |
| 3. | Materials and storage | 3 |
| 4. | Introduction | 4 |
| 5. | Assay principle | 4 |
| 6. | Assay preparation | 5 |
| 7. | Sample preparation | 7 |
| 8. | Assay protocol | 8 |
| 9. | Data analysis | 9 |
| 10. | Troubleshooting | 11 |
| 11. | Additional Information | 13 |
| 12. | Related products | 13 |
| 13. | Warranties and limitation of liability | 14 |



1. General information

PRECAUTIONS

Please read this manual carefully before beginning the assay.

This product is designed for **research use only**. It is not approved for human or animal use or clinical diagnosis. All chemicals should be handled with care and in accordance with laboratory safety practices. It is recommended to use basic Personal Protective Equipment.

Do not use after the expiration date stated on the packaging.

Do not mix or substitute reagents or materials from other kit batches or vendors.

For the **material safety data sheet (MSDS)** please contact us at **info@bioquochem.com**

TECHNICAL RECOMMENDATIONS

Store reagents as indicated in **Materials and storage** section.

Be sure to keep the bottle capped when not in use.

Let the components reach room temperature (RT) before use.

Immediately before use, gently invert and rotate reagent bottles several times to mix the contents thoroughly.

Avoid foaming or bubbles when mixing or reconstituting components.

Avoid cross contamination of samples or reagents by changing pipette tips between sample, standard and reagent additions.

Be sure to use the optimal microplate for the assay. Flat bottom transparent microplates for UV/VIS applications, and black microplates for fluorescence measurements.



2. Technical specifications

Available sizes

100/200/400 tests

O Sample volume required

200 µL/test

Compatible samples

Fruits, vegetables, beverages, herbs, and plant samples

Type of detection

Colorimetric (480 nm)



3. Materials and storage

MATERIALS SUPPLIED

| ltem | No. test | Units | Storage |
|--------------------------------|----------|-------|---------|
| | 100 | 1 | |
| Reagent A | 200 | 2 | RT |
| | 400 | 4 | |
| | 100 | 1 | |
| Reagent B | 200 | 2 | -20 °C |
| | 400 | 4 | |
| | 100 | 1 | |
| Reagent C | 200 | 2 | RT |
| - | 400 | 4 | |
| | 100 | 1 | |
| Standard | 200 | 2 | 4°C |
| | 400 | 4 | |
| | 100 | 1 | |
| Transparent 96-Well Microplate | 200 | 2 | RT |
| | 400 | 4 | |

MATERIALS NEEDED BUT NOT SUPPLIED

- o Double distilled water (ddH2O) as Milli-Q Ultrapure Water.
- Labware materials (micropipettes, tubes, stirring/mixing equipment).
- Colorimetric microplate reader equipped with filter for OD 480 nm.

STORAGE CONDITIONS

On receipt, store kit components as indicated above. Under these conditions, the reagents are stable in the original packaging until the expiration date stated on the outside of the box. After reconstitution, Reagent B and standard solutions are unstable. Prepare a fresh set of solutions for every use.



4. Introduction

Polyphenols are naturally occurring compounds largely found in fruits, vegetables, nuts, seeds, flowers, bark cereals and beverages.

Polyphenols have attracted great interest due to growing evidence of their beneficial effect on human health. They have been reported to exhibit anti-carcinogenic, anti-atherogenic, anti-ulcer, anti-thrombotic, anti-inflammatory, immune modulating, anti-microbial, vasodilatory, and analgesic effects. Polyphenols are also highly demanded compounds for the food industry as natural additives due to their antioxidant, antimicrobial and anti-inflammatory potential.

BQC Phenolic Compounds Assay Kit is a quick, easy, and selective assay to quantify phenolic compounds in a wide variety of samples.

5. Assay principle

BQC Phenolic Compounds Assay Kit is based on the reaction between diazonium salts and phenolic compounds. Under alkaline conditions the diazonium group specifically couples with reactive phenolic hydroxyl groups to form colorimetrically detectable stable azo complexes (λ = 480 nm).

Catechin is used as the reference standard compound for this Phenolic Compounds Assay, and the results are expressed as Catechin Equivalents (CE).

Principle of Phenolic Compounds Assay Kit



6. Assay preparation

REAGENT PREPARATION

All assay reagents not listed below are ready to use as supplied. Allow the reagents to reach room temperature before use.

- **R.B. Working Solution:** Add 2.5 mL of ddH₂O to each vial of Reagent B and mix well.
 - **① CAUTION:** R.B. Working Solution must be freshly prepared and used immediately. Discard the remaining solution.
- **R.C. Working Solution:** Add 2.5 mL of ddH₂O to each vial of Reagent C and mix well.

Standard Solution (Catechin Hydrate): Add 1 mL of Reagent A to the Standard vial and mix well. Dilute this solution 1:10 with ddH₂O (e.g. 100 μ L of standard solution with 900 μ L of ddH₂O). Use this diluted solution to prepare the standard curve.

STANDARD CALIBRATION

Prepare catechin standards for the calibration curve from the 1:10 diluted Standard solution according to the following Table. Prepare the standards immediately prior to each assay. Vortex tubes thoroughly. Discard standard solutions after use.

| Standard No | 1:10 diluted Standard Solution (µL) | ddH ₂ O (μL) | *CE (µM) |
|--------------------------|--|-------------------------|----------|
| Std 1 (Reagent Blank) | 0 | 1000 | 0 |
| Std 2 | 10 | 990 | 10 |
| Std 3 | 25 | 975 | 25 |
| Std 4 | 50 | 950 | 50 |
| Std 5 | 75 | 925 | 75 |
| Std 6 | 100 | 900 | 100 |

^{*}Antioxidant activity is expressed as CE (Catechin Equivalents).



PLATE SET UP

BQC recommends running the standards and samples at least in duplicate (triplicate recommended). There is no specific pattern for using the wells on the plate. A proposed layout of standards (Std) and samples (S) to be measured in duplicate is shown below.

• NOTE: If sample blanks are included in the assay, it is necessary to reserve some wells of the plate for these controls

| Q | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
|---|-----------|------------|-----------|-----------|------------|------------|------------|------------|------------|------------|------------|------------|
| Α | Std 1 | Std 1 | S3 | \$3 | S11 | S11 | \$19 | \$19 | S27 | S27 | \$35 | \$35 |
| В | Std 2 | Std 2 | S4 | S4 | S12 | S12 | \$20 | \$20 | S28 | S28 | \$36 | \$36 |
| С | Std 3 | Std 3 | \$5 | \$5 | \$13 | \$13 | \$21 | \$21 | \$29 | \$29 | S37 | S37 |
| D | Std 4 | Std 4 | S6 | S6 | \$14 | \$14 | S22 | \$22 | \$30 | \$30 | \$38 | \$38 |
| E | Std 5 | Std 5 | S7 | S7 | \$15 | \$15 | \$23 | \$23 | S31 | S31 | S39 | S39 |
| F | Std 6 | Std 6 | S8 | S8 | \$16 | \$16 | S24 | S24 | S32 | S32 | S40 | \$40 |
| G | S1 | S 1 | S9 | S9 | S17 | S17 | \$25 | \$25 | \$33 | \$33 | S41 | S41 |
| Н | S2 | S2 | \$10 | \$10 | \$18 | \$18 | S26 | S26 | \$34 | S34 | S42 | S42 |

Example of plate layout for Phenolic Compounds Assay Kit



7. Sample preparation

The following sample preparation protocols are intended as a guide only. The optimal conditions for sample preparation must be determined by the end user. It is recommended to use fresh samples. If it is not possible, aliquot and store samples appropriately with minimal freeze/thawing.

Phenolic Compounds Assay Kit can be used to determine phenolic content in a wide variety of samples like fruits, vegetables, plants, herbs, and beverages.

Food and beverages. Fruit juices and other beverages such as wine, tea, and coffee can be directly measured with appropriate dilutions. If it is required, clarify the sample through filtration prior performing the assay. Ensure that the selected filter is appropriate for filtering your samples, avoiding polyphenols retention.

For the analysis of other samples like **fruits**, **vegetables**, **and plants** an extraction step is usually required. The extraction method varies based upon the sample type. The most common extraction solvents include acid/methanol, acid/ethanol, or acetone.

Reagents and materials required for sample preparation are not supplied with the kit. Before doing sample preparation, consider the volume of sample required per test; see **Technical specifications** section.

Make sure that interfering substances present in the sample do not give a significant background. Run proper blanks as necessary (e.g. sample blank should be always evaluated when working with highly colored samples). It is recommended to assay different sample dilutions to ensure the values fall within the standard curve.



8. Assay protocol

Prepare and mix all reagents thoroughly before use. Each standard, sample or control should be assayed at least in duplicate.

| 1 | | Set up the plate design |
|---|-----|---|
| 2 | | Add 200 µL of standard or sample in each well |
| 3 | | Add 20 µL of R.B. Working Solution in all wells |
| 4 | | Add 20 µL of R.C. Working Solution in all wells |
| 5 | (1) | Let the reaction run for 10 minutes at RT |
| 6 | | Read the absorbance of all wells at 480 nm in end point mode at RT |

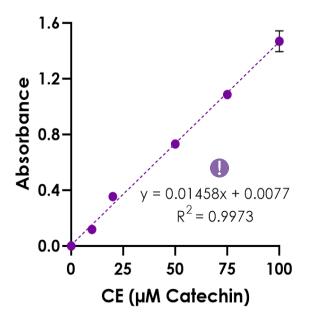
If you need to **adapt this kit** for another form of the assay (for example cuvette), **contact us at info@bioquochem.com**



9. Data analysis

ANALYSIS OF THE STANDARDS

- Calculate the average absorbance of the standards.
- Subtract the average absorbance of the reagent blank (Std 1) from the average absorbance of the remaining standards to obtain the blank-corrected absorbance of the standards.
- Create a standard curve by plotting the blank-corrected absorbance of the standards as a function of the standard concentration (see STANDARD CALIBRATION section). A typical standard curve (y=slope·x ± intercept) for this assay is shown below.



Catechin standard curve with Phenolic Compounds Assay Kit

This standard curve is an example of the data typically obtained with this kit. DO NOT USE this standard curve to calculate the phenolic content of your samples. A new standard curve must be performed by the end user.



ANALYSIS OF THE SAMPLES

- Calculate the average absorbance of the samples.
- Subtract the average absorbance of the reagent blank (Std 1) from the average absorbance of each sample to obtain the blank-corrected absorbance of the samples (As).
- Calculate CE (µM Catechin) from a sample using the following equation. Slope and intercept values are obtained from the standard curve.

CE (
$$\mu$$
M Catechin)= $\left(\frac{A_S - intercept}{slope}\right)$

When working with diluted samples the phenolic concentration (CE, μ M Catechin) obtained must be multiplied by the dilution factor to obtain the CE value of the undiluted sample.



10. Troubleshooting

This troubleshooting table provides potential sources and solutions for common problems observed with BQC Assay Kits. **The problems listed below could occur when using any BQC Assay Kit**. They are not specific for this assay kit.

| Problem | Possible Cause | Recommended Solution |
|--|--|--|
| | Plate read at incorrect wavelength | Check the wavelength used in the assay |
| Wells have color but there is no reading | Incorrect microplate | Use the correct microplate for your application UV/Vis: transparent Fluorescence: black wells/transparent bottom |
| | Pipetting errors in preparation of standards | Avoid pipetting small volumes (<5 µL) Be careful not to splash from well to well |
| | Air bubbles formed in well(s) | Use reverse pipetting technique |
| Standard readings do not | Standard stock is at incorrect concentration | Always refer to dilutions described in Assay preparation |
| follow a linear pattern | Improperly thawed reagents | Thaw all components completely and mix well before use |
| | Use of improperly stored reagents | Store the components appropriately Use fresh components from the standard curve |
| | Incorrect incubation times or temperatures | Refer to Assay protocol |
| Dispersion of standard and sample | Pipetting errors | Avoid pipetting small volumes (<5 µL) Be careful not to splash from well to well |
| readings | Air bubbles formed in well(s) | Use reverse pipetting technique |

Booklet v04

| Problem | Possible Cause | Recommended Solution |
|---|---|--|
| | Samples contain interfering substances | Dilute sample further (if possible) |
| Sample erratic | Inappropriately stored samples or samples used after multiple freeze-thaw cycles | Use fresh samples or store appropriately until use |
| values | Samples not deproteinized | Use an appropriate deproteinization protocol |
| | Cells/Tissue samples not homogenized completely | Repeat the sample homogenization |
| | Inappropriate sample dilution buffer | Refer to Assay preparation |
| Sample reading fall outside the detection range | Samples are too diluted/concentrated No analyte/activity is observed in the sample | Re-assay using different sample dilutions |

STILL HAVING PROBLEMS?

Contact BQC if you have any further questions, our team will be pleased to help you:

| Phone | + 34 985 26 92 92 |
|----------------|---|
| E-mail | info@bioquochem.com |
| Business hours | Monday-Thursday: 8.30 to 17.00 (CEST) Friday: 8.00 to 15.00 (CEST) |



11. Additional Information

BQC Phenolic Compounds Assay Kit is a quick (< 30 minutes) and precise (RSD < 10 %) assay for determining phenolic compounds in a wide variety of samples.

Colored compounds with wavelengths near to 480 nm (e.g. chlorophyll b) have been reported to interfere with this assay.

If unexpected results are obtained running your samples, please contact us at info@bioquochem.com

12. Related products

More products available on bioquochem.com

| Reference | Product |
|-----------|-------------------------------------|
| KB03006 | Polyphenol Quantification Assay Kit |
| KB03015 | Anthocyanins Assay Kit |
| KB03033 | NAD/NADH Quantification Assay Kit |



13. Warranties and limitation of liability

BQC shall not in any event be liable for incidental, consequential or special damages of any kind resulting from any use or failure of the products, even if BQC has been advised of the possibility of such damage including, without limitation, liability for loss of use, loss of work in progress, downtime, loss of revenue or profits, failure to realize savings, loss of products of buyer or other use or any liability of buyer to a third party on account of such loss, or for any labor or any other expense, damage or loss occasioned by such product including personal injury or property damage is caused by BQC's gross negligence. Any and all liability of BQC hereunder shall be limited to the amounts paid by the buyer for the product.

Buyer's exclusive remedy and BQC's sole liability hereunder shall be limited to a refund of the purchase price, or the replacement of all material that does not meet our specifications.

Said refund or replacement is conditioned on buyer giving written notice to BQC within 30 days of shipment.

Expiration date: 1 year from the date of fabrication. Expiration date is indicated on the outside of the box.

For further details, please refer to our website bioquochem.com



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